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Short Communication

Fisetin Protects DNA Against Oxidative Damage and Its Possible Mechanism

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- · 3',4'-dihydroxyl

Abstract

Purpose: The paper tries to assess the protective effect of fisetin against •OH-induced DNA damage, then to investigate the possible mechanism.

Methods: The protective effect was evaluated based on the content of malondialdehyde (MDA). The possible mechanism was analyzed using various antioxidant methods *in vitro*, including •OH scavenging (deoxyribose degradation), •O₂ scavenging (pyrogallol autoxidation), DPPH• scavenging, ABTS•⁺ scavenging, and Cu²⁺-reducing power assays. *Results:* Fisetin increased dose-dependently its protective percentages against •OH-induced DNA damage (IC₅₀ value =1535.00±29.60 μ M). It also increased its radical-scavenging percentages in a dose-dependent manner in various antioxidants assays. Its IC₅₀ values in •OH scavenging, •O₂ scavenging, DPPH• scavenging, ABTS•⁺ scavenging, ABTS•⁺ scavenging, and Cu²⁺-reducing power assays, were 47.41±4.50 μ M, 34.05±0.87 μ M, 9.69±0.53 μ M, 2.43±0.14 μ M, and 1.49±0.16 μ M, respectively.

Conclusion: Fisetin can effectively protect DNA against •OH-induced oxidative damage possibly via reactive oxygen species (ROS) scavenging approach, which is assumed to be hydrogen atom (H•) and/or single electron (e) donation (HAT/SET) pathways. In the HAT pathway, the 3',4'-dihydroxyl moiety in B ring of fisetin is thought to play an important role, because it can be ultimately oxidized to a stable *ortho*-benzoquinone form.

Introduction

Reactive oxygen species (ROS), generated by normal cellular metabolism and exogenous agents (e.g. xenobiotics, ionising and nonionsing radiation), may lead to a condition of oxidative stress if its production overwhelms the antioxidant defences.^{1,2} It has been considered as a significant promoter of cancer, cardiovascular malfunction, aging, and other diseases. Cellular DNA is a particularly sensitive target because of the potential to create cumulative mutations that can disrupt cellular homeostasis. However, the enzymatic repair in living organisms might be inadequate for protecting against permanent DNA mutations. Therefore, for the past few years, it has become a research focus to search for effective and safe antioxidants from natural sources.^{3,4}

Most of natural antioxidants belong to the family of phenolic or polyphenolic compounds, which show their antioxidant activity via a hydrogen atom transfer (HAT) or single electron transfer (SET) mechanism.⁵

Fisetin (3,3',4',7-tetrahydroxyflavone, Figure 1) is a phenolic flavonoid in various food, such as strawberry, onion and persimmon.⁶ It has a wide range of pharmacological effects, such as antitumor, antiinflammation,⁷ anticoagulant and dissolving thrombus.⁸ In recent years, its protective effect on the liver

in human body (especially patients with diabetes) attracted a wide spread attention. Researches showed that, the fisetin's liver protective effect and promoting glucose homeostasis effect were related to its antioxidant activity.⁹ However, its antioxidant ability and mechanism remain unknown. Therefore, the present study used DNA protection model to investigate its antioxidant activity then further discuss the possible mechanisms.



Figure 1. The structure of fisetin (A) and its ball-stick model (B)

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Materials and Methods

Chemicals

Fisetin (98%, CAS 528-48-3) and DNA sodium salt (fish sperm) were obtained from Aladdin Co. (Shanghai, China). DPPH• (1,1-diphenyl-2-picryl-hydrazl), ABTS [2,2'-azino-bis(3-ethylbenzo- thiazoline-6-sulfonic acid diammonium salt)], neocuproine, BHA (butylated hvdroxvanisole). Trolox $[(\pm)-6-hydroxyl-2,5,7,8]$ tetramethlychromane-2-carboxylic acid], pyrogallol and deoxyribise were purchased from Sigma Co. (Sigma-Aldrich Shanghai Trading Co., China). Other chemicals used in this study were of analytical grade and obtained from Guangzhou Chemical Reagent Factory (Guangzhou, China).

Methods

The protective effect of fisetin against •OH-induced DNA damage was determined by our method.¹⁰ Mechanistic analysis experiments included •OH scavenging, $\bullet O_2^-$ scavenging, DPPH• scavenging, ABTS•⁺ scavenging, and Cu²⁺-reducing power assays. The •OH scavenging assay was based on the deoxyribose degradation, and improved by our laboratory;¹¹ in the improved deoxyribose degradation assay, all samples were pre-treated before determination. The $\bullet O_2^-$ scavenging assay was based on pyrogallol autoxidation reaction, and also improved by our laboratory; In the improved pyrogallol autoxidation assay, pH was modified as 7.4.¹¹ The other antioxidant assays were described in our previous paper.^{10,11} In all these assays,

BHA and Trolox were used as the positive controls. The inhibition (or protecting DNA, reducing power) percentages were obtained according to the corresponding calculation formulas. The calculation formulas and experimental protocols were detailed in the Suppl. 1.

Results and Discussion

ROS can lead to the formation of single and doublestrand breaks, as well as induce chemical and structural modifications to purine and pyrimidine bases, resulting day.' millions of lesions per cell each in Deoxythymineglycol (dTG), 8-hydroxy-deoxyguanine malondialdehyde (8-OH-dG), (MDA) and formamidopyrimidine (FAPy) constitute the important markers of DNA oxidative damage.^{12,13} Among these fragments, MDA is usually used to reflect the DNA protective percentage, because MDA can be easily detected via combining with 2-thiobarbituric acid (TBA).¹⁰

As is seen in Figure 2A, the protective percentages of fisetin from •OH-induced DNA damage increased in a dose-dependent fashion. The IC₅₀ values of fisetin, BHA and Trolox were respectively 1535.00±29.60 μ M, 2469.00±96.69 μ M and 1084.67±20.23 μ M (Table 1). The results suggested that fisetin could protect DNA against •OH-induced damage. However, its protective effect is inferior to that of Trolox but stronger than that of BHA.



Figure 2. The dose response curves of fisetin in the assays: (A) protective effect against DNA damage; (B) hydroxyl (•OH) radical-scavenging; (C) superoxide anion (•O₂⁻) radical-scavenging assay; (D) DPPH• radical-scavenging assay; (E) ABTS⁺• radical-scavenging assay; (F) Cu²⁺-reducing power. Each value is expressed as the mean±SD (n=3). The concentration was the final concentration in the corresponding reaction system.

Some studies have shown that phenolic antioxidants protect DNA possibly through base-excision repair, which may arise from ROS (especially •OH and •O₂⁻) scavenging at high reaction rates.¹⁴⁻¹⁶ Therefore, the •OH and •O₂⁻ radical-scavenging capacities of fisetin were further measured in the study.

Our data in Figure 2B&C showed that fisetin could efficiently scavenge •OH and $•O_2^-$ radicals dose-dependently. The IC₅₀ values of fisetin, BHA and Trolox of •OH-scavenging were respectively 47.41±4.50 µM, 2.40±1.19 µM, and 41.84±9.55 µM. In the $•O_2^-$ radical-scavenging assay, IC₅₀ values of fisetin, BHA and Trolox were 34.05±0.87 µM, 76.02±5.22 µM and 66.75±2.25 µM, respectively (Table 1). This indicated that both •OH and $•O_2^-$ radicals could be eliminated by fisetin at low concentration, and that ROS scavenging may be a possible approach for fisetin to protect DNA.

To confirm whether HAT and SET might happen in the ROS scavenging by fisetin, we further measured its radical-scavenging on DPPH•. The previous studies suggested that DPPH• may undergo HAT pathway to be scavenged to yield a stable DPPH-H molecule.¹⁷ As illustrated in Figure 2D and Table 1, fisetin scavenged DPPH• radical with high efficiency. It clearly suggests a HAT pathway in the ROS scavenging by fisetin. The possible reaction of fisetin with DPPH• radical can be proposed as Figure 3.

In the process, since *B* ring is regarded as the active sites in the antioxidant process of flavonoids,¹⁸ phenolic-OH in *B* ring of fisetin is thought to undergo homolysis prior to either the *A* or *C* ring, to produce H• and fisetin• radical (I). Then DPPH-H molecule may be generated through H• combining with DPPH•. And the fisetin• radical might transform into a semi-quinone form (III), which could be further extracted H• by excess DPPH• to form the stable *ortho*-benzoquinone form. Now it is clear that, the 3',4'-dihydroxyl moiety in B ring of fisetin played an important role, because it could be ultimately oxidized to a stable *ortho*-benzoquinone form.



Figure 3. The proposed reaction of fisetin with DPPH• radical.

Since the formation of $ABTS^{\bullet^+}$ from ABTS was previously proven to be via one SET oxidation,¹⁹ $ABTS^{\bullet^+}$ scavenging was also reported to be a SET mechanism.²⁰ The effective scavenging $ABTS^{\bullet^+}$ by fisetin indicate a SET possibility in its ROS scavenging reaction. The SET possibility was further supported by the Cu²⁺-reducing power assay, in which fisetin efficiently reduced Cu²⁺ \rightarrow Cu⁺ (Figure 2F and Table 1). Cu²⁺-reducing however is well-known as an electron (*e*) transfer reaction.

-	Figstin	Positive controls	
	Fiseun	BHA	Trolox
DNA protective effect	1535.00±29.60 ^b	2469.00±96.69c	1084.67±20.23ª
•OH scavenging	47.41±4.50°	2.40 ± 1.19^{a}	41.84±9.55b
•O2 ⁻ scavenging	34.05±0.87ª	76.02±5.22°	66.75±2.25 ^b
DPPH• scavenging	9.69 ± 0.53^{a}	22.84±0.34 ^c	19.55±2.20 ^b
ABTS ⁺ • scavenging	2.43±0.14 ^a	7.73±0.43 ^b	7.54 ± 0.57^{b}
Cu ²⁺ -reducing	1.49 ± 0.16^{a}	3.25 ± 0.17^{b}	4.31±0.21°

Table 1. The IC₅₀ values of fisetin and the positive controls (µM)

The IC₅₀ value is defined as the concentration of 50% radical inhibition (or protection percentage, relative reducing percentage). These IC₅₀ values were calculated by linear regression analysis based on the response curves in Figure 2 and converted from μ g/mL to μ M. Each value in this table is expressed as the *mean±SD* (*n*=3). Mean values with different superscripts (a, b, or c) in the same row are significantly different (*p* < 0.05), while those with same superscripts are not significantly different (*p* < 0.05).

Conclusion

Fisetin can effectively protect DNA against •OH-induced oxidative damage possibly via reactive ROS scavenging approach, which is assumed to be via HAT/SET pathways. In the HAT pathway, the 3',4'-dihydroxyl moiety in B ring of fisetin is thought to play an important role, because it can be ultimately oxidized to a stable *ortho*-benzoquinone form.

Ethical Issues

Not applicable.

Conflict of Interest

Authors declare no conflict of interest in this study.

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